## New Chemical Constituents from Borreria verticillata (Rubiaceae)

by Vinicius F. Moreira<sup>a</sup>), Rodrigo R. Oliveira<sup>a</sup>), Leda Mathias<sup>a</sup>), Raimundo Braz-Filho<sup>b</sup>), and Ivo J. Curcino Vieira<sup>\*a</sup>)

 <sup>a</sup>) Setor de Química de Produtos Naturais, Universidade Estadual do Norte Fluminense Darcy Ribeiro, 28013-602, Campos dos Goytacazes, Rio de Janeiro, Brazil (phone: +55-22-27397046; fax: +55-22-27397248; e-mail: curcino@uenf.br)
<sup>b</sup>) Pesquisador Visitante Emérito-FAPERJ/UENF/UFRRJ

A phytochemical study on *Borreria verticillata* has led to the isolation of two novel simple indole alkaloids, 6-methoxy-4-(3-methylbut-2-en-1-yl)-1*H*-indole, named verticillatine A (1), and 1-(1*H*-indol-6-yl)-3-methylbutan-1-one, named verticillatine B (2), one new iridoid, 6'-O-(2-glyceryl)scandoside methyl ester (3), with the glycerol unit linked to a glucose unit, and two known ones, asperuloside (4) and scandoside methyl ester (5). The structures of these compounds were elucidated on the basis of spectroscopic-data analyses, mainly <sup>1</sup>H- and <sup>13</sup>C-NMR, including 2D experiments (<sup>1</sup>H,<sup>1</sup>H-COSY, NOESY, HMBC, and HMQC), and HR-ESI-MS.

Introduction. - The Rubiaceae family has many pharmaceutically important plants. Borreria verticillata (L.) G. MEY., popularly known as 'Cordão-de-Frade' is a species which occurs over the entire Brazilian territory and is commonly used in traditional folk medicine as an antipyretic and analgesic [1-3]. Previous studies of *B. verticillata* by other research groups showed the presence of terpenoid indole alkaloids [4][5], and we have also reported the structure of a new iridoid named borreriagenin isolated from a MeOH extract of the flowers of B. verticillata [6]. In continuation of our phytochemical investigation of *B. verticillata*, we now report on the structural characterization of two new simple indole alkaloids, 6-methoxy-4-(3-methylbut-2-en-1-yl)-1H-indole, named verticillatine  $A^1$  (1), and 1-(1*H*-indol-6-yl)-3-methylbutan-1-one, named verticillatine  $B^{1}$  (2), and of a new iridoid, 6'-O-(2-glyceryl)scandoside methyl ester<sup>1</sup>) (3), from the MeOH extract of *B. verticillata* roots, isolated besides two known iridoids, asperuloside (4) and scandoside methyl ester (5), a mixture of aliphatic acids, a mixture of tri-Oacylglycerols, sucrose, and a mixture of glucose and sucrose identified after preparation of the corresponding acetyl derivatives. All structures were elucidated by spectroscopic methods, especially by 2D-NMR techniques and mass spectrometry.

**Results and Discussion.** – The phytochemical investigation of the MeOH extract from *B. verticillata* roots by classical chromatographic methods resulted in the isolation of the five compounds 1-5 (*Fig. 1*), together with a mixture of aliphatic acids, a mixture of tri-*O*-acylglycerols, sucrose, and a mixture of glucose and sucrose. The components of these three mixtures (aliphatic acids, tri-*O*-acylglycerols, and glucose +

<sup>1)</sup> Trivial atom numbering; for systematic names, see Exper. Part.

<sup>© 2010</sup> Verlag Helvetica Chimica Acta AG, Zürich



Fig. 1. Compounds isolated from Borreria verticillata

sucrose), sucrose [7], the known iridoids asperuloside (4) [8], and scandoside methyl ester (=( $6\beta$ )-6-hydroxygeniposide; 5) [9] were identified on the basis of 1D and 2D <sup>1</sup>H- and <sup>13</sup>C-NMR and MS data and comparison with the corresponding literature data. The 1D- and 2D-NMR (<sup>1</sup>H and <sup>13</sup>C) and MS data of 1–3 allowed to characterize the two new simple indole alkaloids verticillatine A (1) and verticillatine B (2), and the new iridoid derivative of scandoside methyl ester, 6'-O-(2-glyceryl)scandoside methyl ester (3).

Verticillatine A (1) was obtained as an amorphous powder. Its IR spectrum exhibited a broad band at 3409 cm<sup>-1</sup> typical for an N–H amine group and bands at 1272 and 1238 cm<sup>-1</sup> for a C–O–C (MeO) group, at 2927 and 725 cm<sup>-1</sup> for =CH, and at 1488 cm<sup>-1</sup> typical for C=C of a benzene ring [10][11]. The UV spectrum showed an absorption band at 242 nm (log  $\varepsilon$  1.86). The molecular formula of C<sub>14</sub>H<sub>17</sub>NO was established by the EI-MS ( $M^+$  at m/z 215) and was supported by NMR data. Comparative analysis of  ${}^{1}H$ - and DEPT(135°)- ${}^{13}C$ -NMR spectra of 1 (*Table 1*) allowed to identify resonances for 14 C-atoms, including three Me groups (one linked to the O-atom at  $\delta(C)$  58.14 (MeO) and two linked to a sp<sup>2</sup> C-atom at  $\delta(C)$  25.76 (Me) and 17.92 (Me)), one CH<sub>2</sub> group at  $\delta$ (C) 26.07, five sp<sup>3</sup> CH groups, and five quaternary sp<sup>2</sup> C-atoms, including an O-bearing one. The presence of a MeO group linked to C(6) was confirmed by a cross-peak correlation  $({}^{3}J(H \rightarrow C))$  in the HMBC spectrum (Fig. 2), i.e.,  $\delta(C)$  151.00 (C(6))/ $\delta(H)$  3.85 (s, MeO). The fragment ions at m/z 200  $([M - Me]^+)$  and 184  $([M - MeO]^+)$  in the EI-MS were also used to confirm the presence of the MeO group. The prenyl (= 3-methylbut-2-en-1-yl) unit linked to C(5) was deduced by <sup>1</sup>H-NMR (1D and 2D <sup>1</sup>H, <sup>1</sup>H-COSY) data, *i.e.*, by a d at  $\delta$ (H) 3.85 (J = 7.0 Hz) of the CH<sub>2</sub>(1') group coupled to the olefinic H–C(2') at  $\delta$ (H) 5.24 (br. t, J= 7.0 Hz). Localization of the prenyl unit at C(5) ( $\delta$ (C) 121.40) was confirmed by a

112.						
<b>1</b> <sup>1</sup> )		<b>2</b> <sup>1</sup> )				
$\delta(C)$	$\delta(\mathrm{H})$	$\delta(C)$	$\delta(\mathrm{H})$			
124.77	7.18 (d, J = 2.9)	125.77	7.29 (br. s)			
101.20	6.51 (d, J = 2.9)	104.59	6.68 (br. s)			
128.32	-	138.58	-			
121.40	-	122.94	8.33 (br. s)			
151.00	-	130.53	-			
110.14	6.92 (d, J = 8.7)	122.53	7.89 (dd, J = 8.6, 1.3)			
108.73	7.19 (d, J = 8.7)	111.16	7.43 $(d, J = 8.6)$			
131.57	-	138.58	-			
26.07	3.62 (d, J = 7.0)	200.68	-			
123.39	5.24(t, J = 7.0)	47.73	2.92 (d, J = 6.9)			
130.98	-	25.87	2.34 - 2.37(m)			
17.92	1.84 (br. s)	23.10	1.02 (br. s)			
25.76	1.67 (br. s)	23.10	1.03 (br. s)			
58.14	3.85(s)	-	-			
_	8.02 (br. s)	_	8.33 (br. s)			
	$\begin{array}{c} \underline{1^{1}} \\ \hline \\ \hline \\ \delta(C) \\ 124.77 \\ 101.20 \\ 128.32 \\ 121.40 \\ 151.00 \\ 110.14 \\ 108.73 \\ 131.57 \\ 26.07 \\ 123.39 \\ 130.98 \\ 17.92 \\ 25.76 \\ 58.14 \\ \hline \end{array}$	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $			

Table 1. <sup>13</sup>*C*- and <sup>1</sup>*H*-*NMR* (CDCl<sub>3</sub>) *Data of Verticillatine A* (1) and *Verticillatine B* (2)<sup>a</sup>).  $\delta$  in ppm, *J* in

<sup>a</sup>) Number of H-atoms bound to C-atoms deduced by comparative analysis of {<sup>1</sup>H}- and DEPT-<sup>13</sup>C-NMR spectra;  $\delta$  and J from 1D <sup>1</sup>H-NMR; assignments confirmed by 2D-NMR data (<sup>1</sup>H,<sup>1</sup>H-COSY, HMQC, and HMBC).



Fig. 2. Selected HMBCs  $(H \rightarrow C)$  for verticillatine A (1) and verticillatine B (2)

cross-peak correlation  $({}^{2}J(H \rightarrow C))$  in the HMBC spectrum (*Fig. 2*), *i.e.*,  $\delta(C)$  121.40 (C(5))/ $\delta(H)$  3.62 (J = 7.0 Hz, CH<sub>2</sub>(1')). The complete analysis of the HMBCs (*Fig. 2*) in combination with additional NMR data (*Table 1*) allowed to characterize verticillatine A as a simple indole alkaloid, 6-methoxy-4-(3-methylbut-2-en-1-yl)-1*H*-indole (**1**).

Verticillatine B (2) was obtained as an amorphous powder. Its IR spectrum exhibited a broad band at 3390 cm<sup>-1</sup> typical for an N–H amine group, a band at 1730 cm<sup>-1</sup> for a C=O group, and at 1500 cm<sup>-1</sup> typical for C=C of a benzene ring [10][11]. The UV spectrum showed an absorption band at 242 nm (log  $\varepsilon$  4.13). The molecular formula of C<sub>13</sub>H<sub>15</sub>NO was established by EI-MS ( $M^{+*}$  at m/z 201) in combination with NMR data. The {<sup>1</sup>H}<sup>13</sup>C-NMR spectrum (*Table 1*) combined with DEPT experiments exhibited resonances for 13 C-atoms, including two Me groups at

 $\delta(C)$  23.10, one CH<sub>2</sub> group at  $\delta(C)$  47.73 linked to a C=O group, five CH groups (4 arom. sp<sup>2</sup> and 1 sp<sup>3</sup>), and four sp<sup>2</sup> quaternary C-atoms, including one C=O at  $\delta(C)$  200.68. The presence of the C=O group was inferred by the signal at  $\delta(C)$  200.68 in the <sup>13</sup>C-NMR spectrum (*Table 1*). The position of the C=O group within an isopentyl group linked to C(6) was confirmed by cross-peaks in the HMBC spectrum (*Fig. 2*), *i.e.*,  $\delta(C)$  200.68 (C(1'))/ $\delta(H)$  2.92 (*d*, *J*=6.9 Hz, CH<sub>2</sub>(2')) and 2.34–2.37 (*m*, H–C(3')). The presence of the isopentyl group was corroborrated by the HMBCs  $\delta(C)$  47.73 (C(2'))/ $\delta(H)$  1.03 and 1.02, (2s, Me(5') and Me(4'), resp.). The substitution at the indole moiety was confirmed by the HMBCs  $\delta(C)$  138.58 (C(9))/ $\delta(H)$  8.33 (H–C(5)), 7.89 (*dd*, *J* = 8.6 and 1.3 Hz, H–C(7)), 2.29 (br. s, H–C(2)), and 6.68 (br. s, H–C(3)). The complete analysis of the HMBCs (*Fig. 2*) in combination with additional NMR data (*Table 1*) also allowed the identification of the features of the unprecedented skeleton of verticillatine B, *i.e.*, of the simple indole alkaloid 1-(1*H*-indol-6-yl)-3-methylbutan-1-one (**2**).

The iridoid 6'-O-(2-glyceryl)scandoside methyl ester (**3**) was obtained as an optically active amorphous powder. The IR spectrum showed bands at 3386 (O–H stretching), 2923 (C–H stretching), 1693 (ester C=O), and 1195–1157 and 1080 cm<sup>-1</sup> (stretching C–O) [10][11]. Comparative analysis of the {<sup>1</sup>H}- and DEPT(135°)-<sup>13</sup>C-NMR spectra of **3** (*Table 2*) revealed signals corresponding to 20 C-atoms,

	<b>3</b> <sup>1</sup> )				
	HMQC $(H \rightarrow C)$		HMBC $(H \rightarrow C)$		$\delta(C)$
	$\delta(C)$	$\delta(\mathrm{H})$	$^{2}J$	$^{3}J$	
CH(1)	97.67	5.82(d, J = 5.6)	C(9)	C(3), C(1')	98.26
H-C(3)	152.96	7.70 (s)		C(1), C(5)	153.85
C(4)	110.31	-	C(3), C(5)		110.74
H-C(5)	44.99	3.46 (dd, J = 6.7, 3.0)	C(9)	C(1), C(7)	45.50
H-C(6)	81.21	4.94 - 4.95(m)	C(5), C(7)		83.20
H-C(7)	130.21	6.33 (br. s)		C(9), C(10)	130.06
C(8)	147.21	_	C(7), C(9), C(10)		147.47
H-C(9)	47.00	3.51 (br. $t, J = 5.8$ )	C(5)	C(7)	47.44
$CH_{2}(10)$	60.38	4.80 (br. $d, J = 15.2$ ),		C(7)	60.93
		4.51 (br. $d, J = 15.2$ )			
C(11)	168.69	_		C(3), MeO	170.17
MeO-C(11)	51.26	3.59 (s)			52.07
H - C(1')	101.01	5.38 (d, J = 7.8)	C(2')	C(1)	100.24
H-C(2')	74.78	4.04 - 4.08 (m)	C(3')		74.71
H-C(3')	78.39	4.26 - 4.28(m)	C(2')		77.78
H-C(4')	71.36	4.18 - 4.21 (m)	C(3')		71.45
H-C(5')	78.81	3.96 - 3.99(m)		C(3'), C(4')	78.32
CH <sub>2</sub> (6')	64.79	4.26 - 4.20 (m)		C(2''), C(4')	62.61
CH <sub>2</sub> (1",3")	62.46	4.47 - 4.49(m), 4.26 - 4.29(m)	C(2")		-
H-C(2")	74.04	4.41 (quint., J = 5.4)	C(1")/C(3")	C(6')	-

Table 2. <sup>13</sup>C- and <sup>1</sup>H-NMR (( $D_5$ )pyridine) Data of Iridoids 3 and 5<sup>a</sup>).  $\delta$  in ppm, J in Hz.

<sup>a</sup>) Number of H-atoms bound to C-atoms deduced by comparative analysis of {<sup>1</sup>H}- and DEPT-<sup>13</sup>C-NMR spectra;  $\delta$  and *J* from 1D <sup>1</sup>H-NMR; assignments confirmed by 2D-NMR data (<sup>1</sup>H,<sup>1</sup>H-COSY, HMQC, and HMBC).

corresponding to three nonhydrogenated sp<sup>2</sup> C-atoms (including one COOMe at  $\delta(C)$ ) 168.69), twelve CH groups (eight sp<sup>3</sup> C-atoms linked to an O-atom and two sp<sup>2</sup> C-atom at  $\delta(C)$  152.96 (C(3)) and 130.21 (C(7))), four sp<sup>3</sup> CH<sub>2</sub> groups linked to an O-atom, and one MeO at  $\delta(C)$  51.26/ $\delta(H)$  3.59, characteristic of a MeO group of an ester function. These data, together with the HR-ESI-MS (pos. mode) furnishing the quasi-molecularion peak at m/z 501.15803 ( $[M + Na]^+$ , **3b** in the Scheme) and indicating a molecular mass  $M_r$  of 478, were compatible with a molecular formula  $C_{20}H_{30}O_{13}$ . Another significant positive-ion peak was observed at m/z 427.12106 ( $[M + Na - C_3H_6O_2]^+$ , 5a in the Scheme) corresponding to a fragmentation involving the glyceryl moiety and a Hatom rearrangement (Scheme). All these data suggested a close similarity of 3 with the known iridoid 5, which was identified as part of the structure of 3, after comparative analysis of the {<sup>1</sup>H}- and DEPT-<sup>13</sup>C-NMR data of **5** establishing the number of signals corresponding to quaternary C-atoms and CH, CH<sub>2</sub>, and Me groups. Additional spectral data of 3, mainly 1D- and 2D-NMR (Table 2), the HR-ESI-MS of 5 ([M +Na]<sup>+</sup> at m/z 427.12085, **5a** in the Scheme, indicating a molecular mass  $M_r$  of 404 compatible with a molecular formula  $C_{17}H_{24}O_{11}$  for 5), comparison with similar data described in [8], and the 1D- and 2D-NMR data of 5 were also used to identify the structure of 3.

Scheme. Proposed Fragmentation Mechanisms of **3** by MS/MS of Peak at m/z 501.15803 ( $[M + Na]^+$ ) and Quasimolecular Ion of **5** at m/z 427.12106 ( $[M + Na]^+$ ). Values observed in the HR-ESI-MS of **3** 





Comparison of the <sup>1</sup>H- and <sup>13</sup>C-NMR data of iridoid 3 and 5 (Table 2) indicated that the CH<sub>2</sub>(6')OH group of **5** at  $\delta$ (C) 62.61 ( $\delta$ (H) 3.76 (br. d, J=11.7 Hz) and 3.55 (superimposed with other signals)) was linked to glyceryl (=2-hydroxy-1-(hydroxymethyl)ethyl) in **3**, *i.e.*, CH<sub>2</sub>(1",3") of **3** appeared at  $\delta$ (C) 62.46 and  $\delta$ (H) 4.47 – 4.49 and 4.26–4.29, CH(2'') at  $\delta$ (C) 74.04 and  $\delta$ (H) 4.41, and CH<sub>2</sub>(6') at  $\delta$ (C) 64.79 and  $\delta$ (H) 4.26 – 4.20. The presence of a glycerol unit linked to  $O - CH_2(6')$  of **3** was confirmed by the presence of the HMQCs  $\delta(H)$  4.41 (quint.,  $J = 5.4 \text{ Hz}, H - C(2''))/\delta(C)$  74.04 (C(2'')) and  $\delta(H) 4.47 - 4.49$  and  $4.26 - 4.29 (CH_2(1'',3''))/\delta(C) 62.46 (C(1'',3''))$  and by the HMBC  $\delta(C)$  64.79 (C(6'))/ $\delta(H)$  4.41 (H-C(2")) (Table 2). Other important HMBCs were  $\delta(C)$  62.46 (C(1'',3''))/ $\delta(H)$  4.41 (H-C(2'')) and  $\delta(C)$  74.04 (C(2''))/  $\delta$ (H) 4.47 – 4.49 and 4.26 – 4.29 (CH<sub>2</sub>(1",3")). Moreover, the <sup>1</sup>H,<sup>1</sup>H-COSY plot of **3** was used to establish homonuclear H-atom interactions (geminal and vicinal). The complete analysis of the HMBC spectrum of **3** confirmed the presence of the basic skeleton of scandoside methyl ester (5) with a glycerol unit and allowed the complete <sup>1</sup>H- and <sup>13</sup>C-NMR chemical shift assignments (*Table 2*) of 6'-O-(2-glyceryl)scandoside methyl ester (3).

Acetylation of **3** with pyridine/Ac<sub>2</sub>O 1:2 ( $\nu/\nu$ ) overnight yielded acetyl derivative **3a** confirming the presence of the OH groups in **3**, among others by the  $\Delta\delta(C)$  of the C-atoms of the glucose unit showing the protection due to the  $\gamma$ -effect [7][10] (see *Exper. Part*).

The known iridoid **4** was identified as asperuloside on the basis of spectral data, mainly 1D- and 2D-NMR and ESI-MS, and comparison with values described in [6].

The authors thank the *Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro (FAPERJ-RJ*, Brazil) for a visiting-researcher fellowship and grants, and the *Conselho Nacional de Desenvolvimento Científico (CNPq*-Brazil) for a research fellowship. We are grateful to Prof. *Marcos Eberlin* (IQ-UNICAMP) for the high-resolution mass-spectrometry analysis.

## **Experimental Part**

General. TLC: silica gel (SiO<sub>2</sub>) 60  $F_{254}$ . Column chromatography (CC): SiO<sub>2</sub> 60 (70–230 mesh). M.p.: *Microquímica MQRPF*; uncorrected. Optical rotation: *Perkin-Elmer-343* digital polarimeter. FT-IR Spectra: *FT-IR-8300-Shimadzu* spectrometer; KBr disk;  $\tilde{\nu}$  in cm<sup>-1</sup>. <sup>1</sup>H- and <sup>13</sup>C-NMR Spectra: *Bruker-DRX-500* spectrometer equipped with inverse probes and field gradient, at 500 (<sup>1</sup>H) and 125 (<sup>13</sup>C) MHz; in CDCl<sub>3</sub> or (D<sub>5</sub>)pyridine soln.;  $\delta$  in ppm rel. to Me<sub>4</sub>Si as internal standard, *J* in Hz; 1D spectra were acquired under standard conditions by using a direct detection 5-mm <sup>1</sup>H/<sup>13</sup>C dual probe; standard pulse sequences were used for 2D spectra by using a multinuclear inverse-detection 5-mm probe with field gradient. EI-MS: *Shimadzu-QP5050A* mass spectrometer; at 70 eV; in *m/z* (rel. %). HR-ESI-MS: *VG7070E-HF* mass spectrometer; in *m/z*.

*Plant Material.* The roots of *Borreria verticillata* (L.) G. MEY. were collected in November 2007 at Campos dos Goytacazes City, Rio de Janeiro State, Brazil, and identified by *I. J. C. V.*, Brazil. A voucher specimen was deposited with the Universidade Estadual do Norte Fluminense Darcy Ribeiro, Brazil.

*Extraction and Isolation.* Dried and powdered root (505 g) from *B. verticillata* was extracted with MeOH at r.t., furnishing, after solvent evaporation, 29 g of crude MeOH extract. MeOH Extract was subjected to CC (SiO<sub>2</sub>, gradient MeOH/CH<sub>2</sub>Cl<sub>2</sub>): *Fractions 1–10. Fr. 2* (99 mg) was submitted to prep. TLC (hexane/AcOEt 9:1): verticillatine A (**1**; 13.8 mg). *Fr. 5* (628.7 mg) was subjected to CC (SiO<sub>2</sub>, gradient AcOEt/hexane): verticillatine B (**2**; 9.0 mg). *Fr. 9* (410.9 mg) was subjected to CC (*RP-18*, gradient H<sub>2</sub>O/MeOH): iridoids **3** (70.0 mg), **4** (20.0 mg), and **5** (34.0 mg).

*Verticillatine A* (=6-*Methoxy-4-(3-methylbut-2-en-1-yl)-1*H-*indole*; **1**): Amorphous powder. IR: 3409 (N–H), 1272 and 1238 (C–O–C, MeO), 2927 and 725 (=CH), 1488 (C=C, benzene). <sup>1</sup>H- and <sup>13</sup>C-NMR (500 and 125 MHz, resp., CDCl<sub>3</sub>): *Table 1*. EI-MS: 215 (100,  $M^{++}$ ), 200 (25), 184 (22), 169 (20).

*Verticillatine B* (=1-(1H-Indol-6-yl)-3-methylbutan-1-one; **2**): Amorphous powder. IR (KBr disk): 3390 (N–H), 1730 (C=O), 1500 (C=C, benzene). <sup>1</sup>H- and <sup>13</sup>C-NMR (500 and 125 MHz, resp., CDCl<sub>3</sub>): *Table 1*. EI-MS: 201 (20,  $M^{++}$ ), 159 (15), 144 (100), 116 (25).

6'-O-(2-Glyceryl)scandoside Methyl Ester (= (1S,4aS,5R,7aS)-1,4a,5,7a-Tetrahydro-5-hydroxy-1-{[6-O-[2-hydroxy-1-(hydroxymethyl)ethyl]-β-D-glucoyranosyl]oxy}-7-(hydroxymethyl)cyclopenta[c]pyran-4-carboxylic Acid Methyl Ester; **3**): Amorphous powder.  $[a]_{D}^{2D} = -25.5$  (c = 0.009, MeOH). IR (KBr): 3386 (O-H), 2923 (C-H), 1693 (ester C=O), 1195-1157 and 1080 (C-O). <sup>1</sup>H- and <sup>13</sup>C-NMR (500 and 125 MHz, resp., (D<sub>5</sub>)pyridine): Table 2. HR-ESI-MS (pos. mode): 501.15803 ( $[M + Na]^+$ , see Scheme; calc. 501.15841).

6'-O-(2-Glyceryl)scandoside Methyl Ester Heptaacetate (= (1S,4aS,5R,7aS)-5-(Acetyloxy)-1-{{2,3,4-tri-O-acetyl-6-O-{2-(acetyloxy)-1-[(acetyloxy)methyl]ethyl]-β-D-glucopyranosyl]oxy}-7-[(acetyloxy)-methyl]-1,4a,5,7a-tetrahydrocyclopenta[c]pyran-4-carboxylic Acid Methyl Ester; **3a**): <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>): 94.74 (C(1)); 151.73 (C(3)); 109.16 (C(4)); 39.13 (C(5)); 81.36 (C(6)); 128.73 (C(7)); 142.81 (C(8)); 46.34 (C(9)); 61.14 (C(10)); 166.83 (C(11)); 51.51 (MeO-C(11)); 96.46 (C(1')); 70.71 (C(2')); 72.40 (C(3')); 70.71 (C(4')); 72.43 (C(5')); 61.51 (C(6')); 62.47 (C(1'',3'')); 68.11 (C(2'')); 21.18-22.55 (7 MeC=O)); 170.26 (7 MeC=O).

## REFERENCES

- [1] P. A. de S. P. Neto, M. V. Silva, N. V. C. Campos, Z. Porfírio, L. C. Caetano, Fitoterapia 2002, 73, 529.
- [2] H. Lorenzi, E. J. A. Matos, O. Gomes, 'Plantas Medicinais no Brasil Nativas e Exóticas', Nova Odessa, SP, Instituto Platarum, 2002, pp. 413–414.
- [3] G. Maynart, J. L. Pousset, S. Mboup, F. Denis, C. R. Séances Soc. Biol. Fil. 1980, 174, 925.
- [4] A. M. Baldé, L. A. Pieters, A. Gergely, V. Wray, M. Claeys, A. J. Vlietinck, *Phytochemistry* 1991, 30, 997.
- [5] M. A. Ferreira, C. S. L. Branco, S. J. K. Liwowski, J. Nat. Prod. 1978, 41, 655.
- [6] I. J. C. Vieira, L. Mathias, R. Braz-Filho, J. Schripsema, Org. Lett. 1999, 1, 1169.
- [7] E. Breitmaier, W. Voelter, in 'Carbon-13 NMR Spectroscopy: High-Resolution Methods and Applications in Organic Chemistry and Biochemistry', 3rd edn., VCH, Weinheim, 1987, p. 465.
- [8] J.-N. Peng, X.-Z. Feng, X.-T. Liang, J. Nat. Prod. 1999, 62, 611.
- [9] C. A. Boros, F. R. Stermitz, J. Nat. Prod. 1990, 53, 1055.
- [10] J. B. Lambert, H. F. Shyrvell, D. Lightner, R. G. Cooks, in 'Introduction to Organic Spectroscopy', Macmillan Publishing Company, New York, 1987, p. 511.
- [11] E. Pretsch, P. Bühlmann, C. Affolter, in 'Structure Determination of Organic Compounds: Tables of Spectral Data', Springer-Verlag, Zurich, Switzerland, 2000, p. 245-312.

Received December 18, 2009